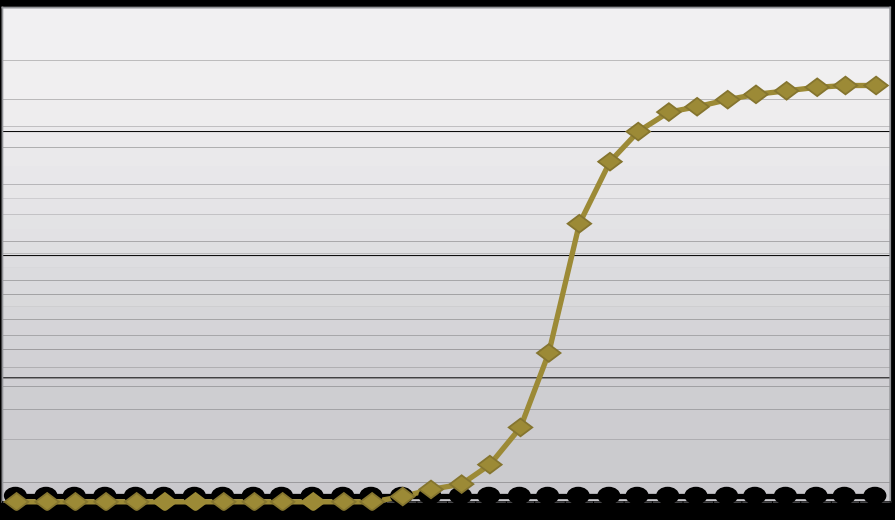


Mycobacterium Paratuberculosis DNA Test Kit, Polymerase Chain Reaction



**For qualitative detection of *Mycobacterium paratuberculosis*
DNA extracted from bovine fecal samples or culture**

NAME AND INTENDED USE

The kit is intended as an aid in the *in vitro* qualitative detection of *Mycobacterium paratuberculosis* DNA by real-time polymerase chain reaction (PCR). The test can be used to detect DNA extracted directly from bovine fecal samples or from culture media used to amplify the agent in bovine fecal samples. The test is intended as an aid in the diagnosis of Johne's disease in cattle.

SUMMARY AND EXPLANATION

Johne's disease is caused by the bacterium *Mycobacterium paratuberculosis*. Johne's is a chronic debilitating disease that is a great source of economic loss for the cattle and dairy industry.

The current reference method for diagnosis is culturing of fecal samples. Culturing is time consuming, taking up to three months for a negative result. Recent advances in real-time PCR chemistry have led to improvements in assay sensitivity that allow for both adequate sensitivity and considerably greater speed in the diagnosis of Johne's disease.

PRINCIPLES OF THE PROCEDURE

The test is based on real-time PCR that, in addition to specific forward and reverse oligonucleotide primers, utilizes fluorogenic probe hydrolysis chemistry to generate a fluorescent signal when specific *M. paratuberculosis* DNA is present in samples. The primers and probe target the *hspX* gene, which contains *M. paratuberculosis*-specific DNA sequences that allow for molecular discrimination of *M. paratuberculosis* from other closely related *Mycobacterium* species.

PRODUCT DESCRIPTION

Mycobacterium Paratuberculosis DNA Test Kit, Polymerase Chain Reaction TC-9828-100

Materials Provided:

Component	Contents
Mastermix (violet capped tubes)	Real-Time PCR Mastermix, containing primers, FAM-labeled probe, and buffers; 4 vials/kit
Taq Polymerase (clear capped tube)	Taq Polymerase Enzyme, 1 vial/kit
Positive Control (green capped tube)	Oligonucleotide, 1 vial/kit
Product Insert	1/kit

Materials and Equipment Required, but Not Provided:

- VetAlert™ MAP Extraction System (Tetracore Cat# TC-9014-100)
- Real-time PCR thermocycler instrument, capable of simultaneously amplifying DNA and detecting a fluorescent signal generated by a TaqMan fluorogenic probe (the instrument should include an optical system which illuminates the reaction vessels and collects the resulting fluorescence emission, a thermal cycler, and data acquisition and analysis software, e.g. Cepheid SmartCycler®, ABI PRISM®, BioRad iCycler iQ®, Roche LightCycler®)
- Reaction tubes or cuvettes (and caps) for the appropriate real-time PCR thermocycler instrument
- Micropipettes and sterile pipet tips with aerosal barriers
- 1x TE (10mM Tris-HCl, pH8.0; 1mM EDTA), for “no template control”
- Microcentrifuge

STORAGE AND STABILITY

The mastermix and *Taq* polymerase should be stored between -15°C and -25°C and are stable until the expiration date stated on the label. Avoid repeated (>2x) thawing and freezing of the mastermix. When working with *Taq* polymerase, keep the enzyme cold (on ice or cooling block) at all times, work quickly, and return enzyme to freezer immediately after use. The positive control should be stored between -15°C and -25°C for longterm storage; working stocks of positive control DNA should be stored between 2° and 8°C for up to one month. Avoid repeated thawing and freezing of positive control DNA (>2x) as this may lead to degradation of the DNA. If the positive control is used only intermittently, it should be frozen in aliquots; each aliquot should be discarded after two freeze/thaw cycles.

PRECAUTIONS

Real-time PCR is an extremely sensitive means for amplifying and detecting small quantities of DNA. Due care should be utilized to prevent any carryover contamination from previous PCR amplifications coming in contact with mastermix and polymerase in this kit.

Recommendations to prevent carryover contamination:

- Store and extract positive or suspected positive material (such as test specimens, positive control, or previously amplified material) separately from all PCR kit components, e.g., in separate rooms.
- Prepare mastermix and *Taq* Polymerase mixture in a BSL-2 cabinet in a DNA-free room. Aliquot mastermix and polymerase mixture into PCR reaction tubes or wells in the same BSL-2 cabinet.
- Add extracted DNA and positive control DNA to reaction tubes or wells in a room separate from the DNA-free room used for mastermix assembly.
- Use sterile pipette tips with aerosol barriers to avoid potential sample-to-sample contamination.
- Periodically (at least weekly), disinfect all work areas and pipettes with either a dilute bleach solution (i.e., 10% Bleach) or ELIMINase® (Decon Labs). If using bleach, the 10% bleach solution should be made up fresh each week. Rinse surfaces with DNase/RNase-free water after decontaminating surfaces.

Thaw mastermix completely before adding *Taq* polymerase. Mix all components by vortex and centrifuge briefly prior to pipetting into PCR reaction tubes, wells, or capillaries.

Keep *Taq* polymerase cold (on ice or cooling block) at all times, work quickly, and return enzyme to freezer immediately after use.

LIMITATIONS

This test should be used for testing DNA extracted from bovine fecal samples or for confirmation of cultures derived from bovine samples. No claims are made for other samples or for organisms derived from other animal species.

SPECIMEN COLLECTION AND STORAGE

Fresh, moist fecal samples should be collected. Upon receipt in the testing laboratory, the fecal samples should be processed immediately; refrigerated for up to 1 day and then processed; or aliquoted, frozen and stored at -80°C. Repeated freezing and thawing of fecal samples should be avoided, as DNA may degrade. Dry fecal samples can produce sub-optimal results and should be avoided.

EXTRACTION OF DNA FROM FECAL SPECIMENS OR CULTURE MEDIA

DNA should be extracted from fecal specimens or culture material using the *VetAlert™* MAP Extraction System (Tetracore, Cat. No. TC-9014-100). The use of other DNA extraction protocols or commercial products should be validated by the user.

POSITIVE AND NEGATIVE CONTROLS

Positive Control:

A positive control is included in the kit and must be included with each test run. The positive control is a non-infectious synthetic template comprising a portion of the target gene sequence of the *hspX* gene of *M. paratuberculosis*. The positive control is supplied as a liquid solution containing 25,000 gene copies/2.5µl.

Negative control:

Negative (i.e., no template) controls must be included with each test run. A negative control is not included in the kit. However, the user is advised to use 1xTE as a no-template control (NTC).

A minimum of one positive and one negative control should be included in each test run.

REAL-TIME PCR TEST PROCEDURE

1. Determine the number of reactions needed.
[NOTE: To ensure that there is ample mix, an additional reaction should be added to the number of reactions required. For example, if 10 reactions are needed, 11 reactions should be prepared.]
2. Remove mastermix vial(s) from the freezer and thaw. After the mastermix has thawed, vortex, and then briefly spin the vial(s) in a microcentrifuge before continuing.
3. For each reaction needed, remove 22.5µL from the mastermix vial and transfer to a new microcentrifuge tube (not provided).
[For example, to perform 10 reactions remove and transfer 225µL.]
4. Remove the *Taq* polymerase from the freezer and quickly centrifuge the tube.
[NOTE: Keep *Taq* polymerase cold (on ice or cooling block) at all times, work quickly, and return to freezer immediately after use.]
5. Add 0.32µL *Taq* polymerase per reaction to the mastermix.
[For example, for 10 reactions add 3.2µL. Due to accuracy issues in pipetting small quantities, a minimum of 10 reactions of mastermix and polymerase should be assembled at any given time.]
6. Cap the microcentrifuge tube and vortex to mix. Briefly spin the tube in a microcentrifuge in order to recover all of the mixture.
7. Add 22.5µL of the mastermix and polymerase mixture to each reaction tube or well.
[NOTE: For LightCycler (Roche) only: add 17.5 µL of the mixture to LightCycler capillaries.]
8. Add 2.5µL of each extracted sample, no template control (not provided), or positive control to the appropriate reaction tube.
9. Close or cover each reaction tube or plate.
10. Briefly centrifuge reaction vessel.

If using SmartCycler® tubes (Cepheid), briefly centrifuge each tube in the Cepheid microcentrifuge provided with the thermocycler, to force the mix to the bottom of the tube.

If using LightCycler® capillaries, spin in microcentrifuge to force the mix to the bottom of the capillary.

In the case of the ABI PRISM® or iCycler instrument, briefly spin each plate to force the mix to the bottom of the tube or well.

In all cases, observe the tube or bottom of the plate to ensure that there is no trapped air, which can interfere with the reaction.

11. Load the tubes, plates or capillaries in the appropriate thermocycling instrument.
12. After an initial enzyme activation step at 95°C, a two step (95°C, 62°C) cycling reaction should be used. Cycling conditions on specific thermocycling instruments are given below (see Cycling Conditions)

CYCLING CONDITIONS

(A) Cepheid SmartCycler™

Cycling Program:

1. Enzyme activation step: 10 seconds @ 95°C (optics off)
2. Two step PCR: [95°C x 5 seconds (optics off), 62°C x 30 seconds (optics on)] for 50 cycles.

Settings for SmartCycler®:

Version I- FTTR25-Dye Set, 25µL reaction with channel 1 as FAM dye

Version II- FTTC25-Dye Set, 25µL reaction with channel 1 as FAM dye,

Read results in FAM dye layer (i.e., channel 1).

(B) ABI PRISM® and BioRad iCycler iQ®

Cycling Program:

1. Enzyme activation step: 10 minutes @ 95°C
2. Two step PCR: (95°C x 15 seconds, 62°C x 60 seconds) for 45 cycles.

ABI PRISM® Settings

- Set the quencher to “NONE”.
- Select the reference dye as “ROX”.
- Select “FAM” as dye layer.
- Plate exposure time should be at (10-25ms, instrument dependent)
- Reaction volume should be set to 25µL.

(C) Roche LightCycler®

Cycling Program:

1. Enzyme activation step: 10 seconds @ 95°C
2. Two step PCR: (95°C x 5 seconds, 62°C x 30 seconds) for 50 cycles.

LightCycler® Settings

- Initial denaturation step should set analysis mode to “NONE”
- Acquisition Mode = NONE
- For the 2-step cycling, analysis mode should be set to “Quantification”
- For the 95°C step, Acquisition Mode = NONE
- For the 62°C step, Acquisition Mode = SINGLE

INTERPRETATION OF TEST RESULTS

The test is a fluorogenic probe hydrolysis assay that produces a characteristic fluorescent signal with a positive test. A positive sample is visualized as a sigmoidal curve on a two-dimensional linear grid where the X-axis represents the PCR cycle number and the Y-axis represents the relative fluorescence of the signal (see representative positive signals in Figure 1). The threshold cycle, or Ct value, is the cycle number at which the increase in the fluorescent signal associated with exponential growth of PCR product is first detected. Ct represents the point at which the signal exceeds the detection threshold and is dependent on the starting template copy number, i.e., the lower the Ct value, the more DNA template was originally present in the test sample at the initiation of the PCR reaction.

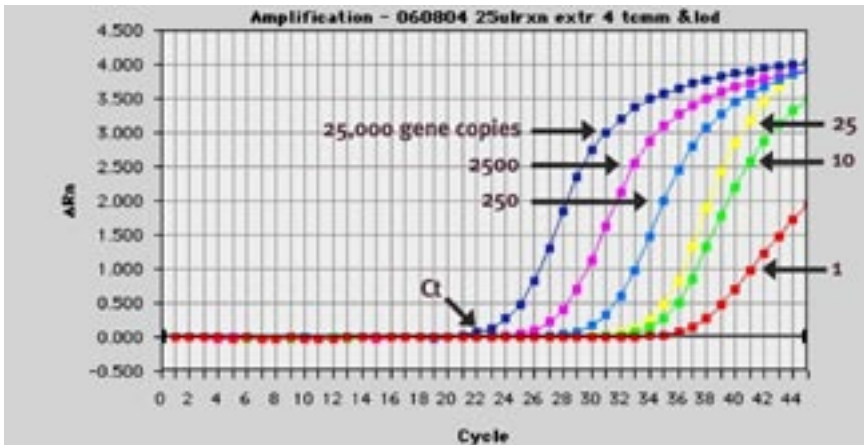


Figure 1. Amplification and detection of *M. paratuberculosis* DNA positive control on ABI PRISM® 7700; positive control was diluted from 25,000 gene copies to 1 gene copy.

A run is considered valid if:

- The amplification curve of the positive control provided (25,000 gene copies / 2.5 μ L) must have a cycle threshold value (Ct) that is between 20-26 cycles.
- The no-template control (NTC) must not cross the threshold prior to the endpoint of the test.

Ct cutoff value for positive samples:

A sample is considered positive if:

- $Ct \leq 38$ for ABI and BioRad series of real-time PCR thermocyclers
- $Ct \leq 42$ for Cepheid and Roche real-time PCR thermocyclers

The Ct for a positive result has been set at the upper range of the Ct values at which one gene copy of DNA can be detected (in an uninhibited sample).

A sample that appears to cross the threshold after the cutoff Ct value can not definitely be considered positive (“suspect” result). Such a sample may represent either a true negative or a low positive which was inefficiently amplified. To confirm such a suspect sample, the test should be repeated in duplicate, taking care to minimize any potential contamination. If at least one out of the two re-tested replicates is positive at an equal or earlier Ct value, the suspect sample can be classified as a “low positive.” Low positive samples should be confirmed by culture.

NOTE: Caution should be used in interpreting positive samples at or below the cutoff value, since such results might be the result of transient passage of consumed organisms rather than a low-level infection.

VALIDATION STUDY

Frozen archived fecal samples from 221 cows were evaluated in a blind trial comparing test results from the Mycobacterium Paratuberculosis DNA Test Kit, Polymerase Chain Reaction test with culture on HEYM (16 weeks). The fecal samples included 100 samples from non-infected cows (test negative, level 4 herd) and 121 culture positive samples of varying intensities. All samples (100/100) from the non-infected cows correctly gave a negative result in the test. The kit gave a positive test result in 59/60 cases when samples had an average cfu/tube count of greater than 3.0. When samples contained less than 3.0 cfu/tube, the kit gave a positive test result in 45/61 samples.

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The use of the *hspX* gene for diagnostic detection of *M. paratuberculosis* is covered by US Patent No. 5,985,576, under exclusive license to Tetracore, Inc. from the USDA.

THE PURCHASE OF THIS PRODUCT ALLOWS THE PURCHASER TO USE IT FOR AMPLIFICATION AND DETECTION OF NUCLEIC ACID SEQUENCES FOR PROVIDING VETERINARY IN VITRO DIAGNOSTICS. NO GENERAL PATENT OR OTHER LICENSE OF ANY KIND OTHER THAN THIS SPECIFIC RIGHT OF USE FROM PURCHASE IS GRANTED HEREBY.

